Journal of Personalized NanoMedicine



Review _

Measurement of Specific Loss Power from Intracellular Magnetic Nanoparticles for Hyperthermia

Asahi Tomitaka¹ and Yasushi Takemura^{2*}

¹Department of Immunology, Herbert Wertheim College of Medicine, Florida International University, Miami, FL 33199, USA

²Department of Electrical and Computer Engineering, Yokohama National University, Yokohama 240-8501, Japan

*Corresponding author: takemura@ynu.ac.jp (Y.T); Tel: +81-45-339-4154

Publication Information

Receved date: 8 August 2015; Accepeted date: 14 September 2015; Published online: 20 September 2015.

Citation: Tomitaka A, Takemura Y. Measurement of Specific Loss Power from Intracellular Magnetic Nanoparticles for Hyperthermia. J Personali NanoMed, 1(1): 33-37

Copyright: ©2015 OLOGY Group.

The articles published in this journal are licensed under a Creative Commons Attribution 4.0 International License.

Abstract

Magnetic nanoparticles have been studied extensively for biomedical applications over the past decades. One of the promising applications of magnetic nanoparticles is hyperthermia which is heat treatment for cancer. For hyperthermia application, it is essential to develop magnetic nanoparticles with high heating efficiency and optimize the condition of magnetic field to achieve adequate heat at tumor. Here, we discuss the heating mechanism of magnetic nanoparticles, influence of intracellular environment for magnetic behavior and heat dissipation, and recent advance on heating evaluation.

Keywords: Magnetic nanoparticles; Magnetic hyperthermia; Relaxation losses

Introduction

Nanomedicine which uses nanotechnology for diagnosis, prevention, and treatment has been growing rapidly for the last few decades. Various nanomaterials have been developed for the field of nanomedicine such as liposomes [1,2], polymeric nanoparticles [3], Micelles [4], dendrimers [5], magnetic nanoparticles [6,7], and gold nanoparticles [8]. Among those nanoparticles, magnetic nanoparticles have been attracting attention due to their unique magnetic properties. Large microscopic field gradients induced by magnetic nanoparticles under magnetic field causes shortening of longitudinal relaxation time (T_1) and transverse relaxation time (T_2) of water proton [9]. This property makes magnetic nanoparticles strong contrast agents for magnetic resonance imaging (MRI) [10,11]. Some of the contrast agents using iron oxide nanoparticles (Fe₂O₄) or γ-Fe₂O₂), which are the most commonly used magnetic nanoparticles for biomedical applications, have been approved for clinical use [12]. Magnetic guidance ability of magnetic nanoparticles can be used for drug delivery [13,14] and gene transfection [15]. Drug delivery to specific targeted site reduces side effect from non-specific distribution. Moreover, magnetic nanoparticles can be used as heat source for hyperthermia (heat treatment) due to heat generation under AC magnetic field [16,17]. Capability for both of diagnosis and treatment makes magnetic nanoparticles promising material for nanomedicine.

Hyperthermia

It has been well known that temperature rise in the range of 40–47°C kills cells in a time and temperature dependent manner [18]. Hyperthermia is heat treatment of cancer based on this temperature sensitivity of cells. In addition to the heat sensitivity,

hypoxic cells (poorly oxygenated cells) in tumor shows higher sensitivity to heat compared to cells in healthy tissue [19]. If the temperature is well controlled at tumor site, hyperthermia would cure cancer without causing damage to surrounding healthy tissues. These concepts make hyperthermia attractive compared to the traditional cancer treatment which cause side effect. Combined treatment of hyperthermia and traditional cancer treatment such as chemotherapy and radiotherapy has been reported to increase treatment effect [19].

Various heating methods have been developed such as water bath for whole body hyperthermia, and using microwave, radio frequency, and ultrasound as external energy sources for regional hyperthermia treatment [20]. However, regional hyperthermia using external heat source has limitation on penetration which prevent treatment for deep-seated tumors, and unavoidable heating of healthy tissue between heat source and tumor is the drawback. Magnetic nanoparticles have been a great candidate as a heat source for local hyperthermia treatment due to their ability for magnetic targeting and heat induction under AC magnetic field at specific region. Hyperthermia using magnetic materials was first proposed in 1957, Gilchrist et al. reported heating of various tissue samples with γ-Fe₂O₃ under AC magnetic field [21]. Since then, hyperthermia using magnetic nanoparticles (called magnetic hyperthermia or magnetic fluid hyperthermia) has been investigated extensively including heating mechanism [22], heating efficiency [23], and in vitro and in vivo study [24,25]. Furthermore, clinical trials of magnetic hyperthermia have been conducted in Germany [26].

Heating Mechanism of Magnetic Hyperthermia

Heat dissipation of magnetic nanoparticles under AC magnetic field is caused by two processes, hysteresis loss and relaxation

losses. When an external magnetic field is applied, magnetic materials possess hysteresis property between magnetic field and magnetization. Hysteresis loss is induced by this hysteresis property, and its value is given by its frequency multiplied by the area of the hysteresis loop in an external field [27]. For small magnetic nanoparticles, relaxation losses are dominant for heat dissipation of the nanoparticles. Relaxation losses are caused by a delay in magnetic moment relaxation. Brownian relaxation loss and Néel relaxation loss are associated with magnetic moment rotation of the entire particle and within a particle, respectively [28]. The Brownian relaxation time τ_B and Néel relaxation time τ_R are given by the following *equations*:

$$\tau_{\rm B} = \frac{3\eta V_{\rm H}}{k_{\rm B}T}$$

$$\tau_{\rm N} = \tau_0 \exp(\frac{KVm}{k_{\rm B}T})$$
(2)

where η is the viscosity of the suspended fluid, V_H is the hydrodynamic volume of magnetic nanoparticles, k_B is the Boltzmann constant 1.38×10^{-23} J/K, T is the temperature in Kelvin, τ_0 is the attempt time of $\sim10^{-9}$ s, K is the magnetocrystalline anisotropy constant, and VM is the volume of the magnetic core. Brownian and Néel relaxations occur in parallel. Thus, effective relaxation time is described as

$$\frac{1}{\tau} = \frac{1}{\tau_{\rm B}} + \frac{1}{\tau_{\rm N}}$$
 (3)

The heat dissipation by relaxation loss is given by the following equation [28]:

$$P_{\text{relax}} = \pi \mu_0 \chi_0 H_0^2 f \frac{2\pi f \tau}{1 + (2\pi f \tau)^2}$$
 (4)

where μ_0 is the permeability of free space, χ_0 is the initial susceptibility, H is the amplitude of the magnetic field, f is the frequency of the magnetic field, and τ is the relaxation time. As shown in equation (1) and (2), Brownian and Néel relaxation times are dependent on viscosity of surrounding environment, hydrodynamic size and core size of magnetic nanoparticles, and their magnetocrystalline anisotropy.

Heating efficiency of magnetic nanoparticles is evaluated by calculating the specific loss power (SLP) or Intrinsic loss power (ILP). SLP provides power dissipation per unit mass of magnetic nanoparticles. Since SLP varies with the amplitude and frequency of applying magnetic field, ILP is introduced to evaluate heating efficiency of magnetic nanoparticles system-independently [29]. SLP and ILP are calculated as

$$SLP = c \frac{m_{\text{sample}}}{m_{MNPs}} \frac{dT}{dt}$$
 (5)

$$ILP = \frac{SLP}{H^2 f} \qquad (6)$$

where c is the heat capacity of water, msample is the mass of the sample, m_{MNPs} is the mass of the magnetic nanoparticles in the sample, and dT/dt is the slope of the temperature rise [30][29].

Although SLP and ILP have been used to evaluate heating efficiency of magnetic nanoparticles, inaccuracy of temperature measurement and lack of consistent measurement system prevent accurate evaluation. As discussed in Wildeboer's report, there is a need for a standardized measurement method for magnetic hyperthermia [31]. In addition to the measurement system, the condition of samples such as volume and shape is limited for the temperature measurement under magnetic field with high amplitude and frequency, and the temperature range is also limited from room temperature to below 100 °C. There is a report showing that initial temperature slope which is used to calculate SLP varied with many factors such as sample volume and temperature sensor position [32].

To overcome these limitations, we proposed a novel way to evaluate heat dissipation of magnetic nanoparticles using AC hysteresis measurement. This measurement gives us dynamic magnetization curve which includes both of hysteresis loss and time lag of magnetization (relaxation losses) from magnetic nanoparticles. The heat dissipation can be directly calculated from the area of this dynamic magnetization curve called AC hysteresis loop. The correspondence between SLP calculated from AC hysteresis loop and temperature rise has been reported [33]. In addition to the reliability of measurement, this method shows magnetic behavior under AC magnetic field and can be used under wide range of amplitude and frequency.

Improvement of Heating Efficiency

The challenge lying in front of magnetic hyperthermia is delivery of magnetic nanoparticles and adequate heat generation under AC magnetic field which is safe for human body. Although delivery of magnetic nanoparticles can be improved using magnet, the magnetic guidance lacks site specificity. To achieve site specific targeting, localization of magnetic nanorods using pulsed magnetic field has been reported [34]. Ligand conjugation on the surface of nanoparticles can be introduced to enhance active targeting to the specific site which expresses receptor [1,35].

Due to the limitation on the quantity of magnetic nanoparticles and magnetic field, it is essential to develop magnetic nanoparticles with high heating efficiency. As shown in equations (1)-(4), heat dissipation from relaxation losses depends on the viscosity of surrounding environment, hydrodynamic size and core size of magnetic nanoparticles, and their magnetocrystalline anisotropy. Tuning magnetocrystalline anisotropy is one of the ways to improve heat dissipation of magnetic nanoparticles. Lee et al developed magnetic nanoparticles with optimal range of magnetocrystalline anisotropy using core—shell structure [36]. Since magnetocrystalline anisotropy is intrinsic material

property, they introduced core-shell structure with $CoFe_2O_4$ in the core and $MnFe_2O_4$ in the shell to tune magnetocrystalline anisotropy by interfacial exchange interaction between hard and soft magnetic phases. Significant enhancement in heat dissipation has been reported for the core-shell nanoparticles compared to traditional magnetic nanoparticles.

It is known that magnetic behavior of magnetic nanoparticles changes with the surrounding environment [37]. As shown in equation (1), Brownian relaxation time depends on the viscosity of suspended fluid and hydrodynamic size of nanoparticles. It has been discussed repeatedly whether particles are free to move or immobilized in tumor tissue or inside cells after delivery to the target site or cellular internalization. Latter case limits heat generation from Brownian relaxation due to the inhibition of Brownian motion (Figure 1). Dutz et al reported static magnetic properties of magnetic nanoparticles injected into mice tumor [38]. Immobilization of magnetic nanoparticles in tumor was observed from the increase in coercivity and remanent magnetization of magnetic nanoparticles after injection into tumor. Similar tendency was observed for magnetic nanoparticles immobilized by gelatin compared to the nanoparticles suspended in fluid. Fortin et al reported temperature rise of magnetic nanoparticles which are internalized into human prostatic adenocarcinoma cells (PC3) [39]. The cellular internalized magnetic nanoparticles showed lower temperature rise compared to the same nanoparticles dispersed into water. Corato et al reported SLP of magnetic nanoparticles in water, on the cell membrane before internalization, and after internalization into human adenocarcinoma SKOV-3 cells [40]. Rapid fall in SLP value was observed after association of magnetic nanoparticles with the cell membrane and internalization.

AC hysteresis measurement is beneficial for understanding magnetic behavior of magnetic nanoparticles under AC magnetic field as well as estimating heat dissipation. We observed AC hysteresis loops of magnetic nanoparticles with the core diameter of 10 nm prepared in three different states; liquid sample which is dispersed in water, immobilized sample fixed with an epoxy bond, and cellular sample internalized into HeLa cells, and estimated specific loss power (SLP) [41]. Immobilized nanoparticles and cellular internalized nanoparticles showed lower SLP compared to liquid sample (Figure 2). This indicates that rotations of nanoparticles are inhibited inside the cells. Slightly lower SLP of cellular sample compared to immobilized sample suggests that rotations of magnetic moments are also inhibited by dipole-dipole interactions inside cells due to the partial agglomeration of magnetic nanoparticles [42]. We also estimated specific loss power (SLP) of magnetic nanoparticles in a single cell [41].

Magnetic properties of magnetic nanoparticles and the conditions to optimize their heating efficiency are well understood when they are magnetically independent [43]. However, previous reports suggest that both of blocking of Brownian motion from cell-particle interaction and magnetic interactions between each particle due to dense population of nanoparticles can affect to heat dissipation of magnetic nanoparticles inside cells [39-42]. Therefore, it is essential to understand the magnetic properties

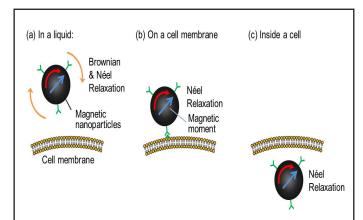


Figure 1. Schematic illustrations of magnetic nanoparticle behavior in a liquid (a), on a cell membrane (b), and inside a cell (c) under AC magnetic field. In a liquid, magnetic nanoparticles are expected to induce heat by both of Brownian and Néel relaxation losses. Only Néel relaxation loss is expected to contribute to heat dissipation for magnetic nanoparticles immobilized on a cell membrane or inside a cell.

and heating mechanisms of magnetic nanoparticles within intracellular or intratumoral environment and under magnetic interactions to develop optimal magnetic nanoparticles for hyperthermia.

Conclusion

There has been a significant improvement in theoretical study of magnetic relaxation and relaxation losses in the past few years [44,45]. However, inaccurate temperature measurement and lack of consistent measurement system make it difficult to give accurate value of SAR and ILP in experimental study [31]. As an alternative method, AC hysteresis loop measurement was suggested to evaluate SAR and ILP without measuring actual temperature rise. In addition, theoretical study of magnetic hyperthermia in cellular environment model has been

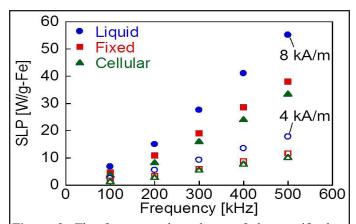


Figure 2. The frequency dependence of the specific loss power (SLP) estimated from the areas of AC hysteresis loops. Magnetic nanoparticles was prepared in liquid (circle), fixed with an epoxy bond (square), and internalized into HeLa cells (triangle). The amplitude of the magnetic field was 4 kA/m (open) or 8 kA/m (filled).

reported [46]. Since heat dissipation of magnetic nanoparticles varies with surrounding environment, SAR evaluation needs to be done in models such as intracellular and intratumoral environment. We also demonstrated that relaxation losses of intracellular nanoparticles can be calculated using AC hysteresis measurement [41].

Acknowledgement

This work was partially supported by JSPS KAKENHI Grant Number 26289124.

References

- 1. Zhang L, Zhou H, Belzile O, Thorpe P, Zhao D. Phosphatidylserine-targeted bimodal liposomal nanoparticles for *in vivo* imaging of breast cancer in mice. J Control Release. 2014; 183: 114–123.
- 2. Al-Ahmady ZS, Chaloin O, Kostarelos K. Monoclonal antibody-targeted, temperature-sensitive liposomes: *In vivo*. J Control Release. 2014; 196: 332–343.
- 3. Zhang X, Dong Y, Zeng X, Liang X, Li X, Tao W, et al. The effect of autophagy inhibitors on drug delivery using biodegradable polymer nanoparticles in cancer treatment. Biomaterials. 2014; 35: 1932–1943.
- 4. Xu H, Yao Q, Cai C, Gou J, Zhang Y, Zhong H, et al. Amphiphilic poly(amino acid) based micelles applied to drug delivery: The *in vitro* and *in vivo* challenges and the corresponding potential strategies. J Control Release. 2015; 199: 84–97.
- 5. Kesharwani P, Jain K, Jain NK. Dendrimer as nanocarrier for drug delivery. Prog Polym Sci. 2014; 39: 268–307.
- 6. Ding H, Sagar V, Agudelo M, Pilakka Kanthikeel S, Atluri VSR, Raymond A, et al. Enhanced blood-brain barrier transmigration using a novel transferrin embedded fluorescent magnetoliposomes nanoformulation. Nanotechnology. 2014; 25: 055101.
- 7. Arami H, Khandhar AP, Tomitaka A, Yu E, Goodwill PW, Conolly SM, et al. *In vivo* multimodal magnetic particle imaging (MPI) with tailored magneto/optical contrast agents. Biomaterials. 2015; 52: 251–261.
- 8. Park J, Park J, Ju EJ, Park SS, Choi J, Lee JH, et al. Multifunctional hollow gold nanoparticles designed for triple combination therapy and CT imaging. J Control Release. 2015; 207: 77–85.
- 9. Huang J, Zhong X, Wang L, Yang L, Mao H. Improving the Magnetic Resonance Imaging Contrast and Detection Methods with Engineered Magnetic Nanoparticles. Theranostics. 2012; 2: 86–102
- 10. Tomitaka A, Jo J, Aoki I, Tabata Y. Preparation of biodegradable iron oxide nanoparticles with gelatin for magnetic resonance imaging. Inflamm Regen. 2014; 34: 45–55.
- 11. Abakumov MA, Nukolova NV, Sokolsky-Papkov M, Shein SA, Sandalova TO, Vishwasrao HM, et al. VEGF-targeted

- magnetic nanoparticles for MRI visualization of brain tumor. Nanomedicine 2015; 11: 825–833.
- 12. Wang YX. Superparamagnetic iron oxide based MRI contrast agents: current status of clinical application. Quant Imaging Med Surg. 2011; 1: 35–40.
- 13. Chen J, Shi M, Liu P, Ko A, Zhong W, Liao WJ, et al. Reducible polyamidoamine-magnetic iron oxide self-assembled nanoparticles for doxorubicin delivery. Biomaterials. 2014; 35:1240–1248.
- 14. Ye F, Barrefelt A, Asem H, Abedi-Valugerdi M, El-Serafi I, Saghafian M, et al. Biodegradable polymeric vesicles containing magnetic nanoparticles, quantum dots and anticancer drugs for drug delivery and imaging. Biomaterials. 2014; 35: 3885–3894.
- 15. Kami D, Kitani T, Kishida T, Mazda O, Toyoda M, Tomitaka A, et al. Pleiotropic functions of magnetic nanoparticles for ex vivo gene transfer. Nanomedicine. 2014; 10: 1165–1174.
- 16. Tomitaka A, Ueda K, Yamada T, Takemura Y. Heat dissipation and magnetic properties of surface-coated ${\rm Fe_3O_4}$ nanoparticles for biomedical applications. J Magn Magn Mater. 2012; 324: 3437–3442.
- 17. Pala K, Serwotka A, Jeleń F, Jakimowicz P, Otlewski J. Tumorspecific hyperthermia with aptamer-tagged superparamagnetic nanoparticles. Int J Nanomedicine. 2014; 9:67–76.
- 18. Roti Roti JL. Cellular responses to hyperthermia (40–46 °C): Cell killing and molecular events. Int J Hyperthermia. 2008; 24: 3–15.
- 19. Hofer KG. Hyperthermia and cancer. Eur Cell Mater. 2002; 3: 67–69.
- 20. Ohguri T, Yahara K, Moon SD, Yamaguchi S, Imada H, Terashima H, et al. Deep regional hyperthermia for the whole thoracic region using 8 MHz radiofrequency-capacitive heating device: Relationship between the radiofrequency-output power and the intra-oesophageal temperature and predictive factors for a good heating in 59 patients. Int J Hyperthermia. 2011; 27: 20–26.
- 21. Gilchrist RK, Medal R, Shorey WD, Hanselman RC, Parrott JC, Taylor CB. Selective inductive heating of lymph nodes. Ann Surg. 1957; 146: 596–606.
- 22. Vallejo-Fernandez G, Whear O, Roca AG, Hussain S, Timmis J, Patel V, O'Grady K. Mechanisms of hyperthermia in magnetic nanoparticles. J Phys D Appl Phys. 2013; 46: 312001.
- 23. Jeun M, Lee S, Kang JK, Tomitaka A, Kang KW, Kim YI, et al. Physical limits of pure superparamagnetic Fe_3O_4 nanoparticles for a local hyperthermia agent in Nanomedicine. Appl Phys Lett. 2012; 100: 092406.
- 24. Gao F, Cai Y, Zhou J, Xie X, Ouyang W, Zhang Y, et al. Pullulan Acetate Coated Magnetite Nanoparticles for Hyperthermia: Preparation, Characterization and *In Vitro* Experiments. Nano Res. 2010; 3:23–31.
- 25. Sadhukha T, Wiedmann TS, Panyam J. Inhalable magnetic

- nanoparticles for targeted hyperthermia in lung cancer therapy. Biomaterials. 2013; 34: 5163–5171.
- 26. Thiesen B, Jordan A. Clinical applications of magnetic nanoparticles for hyperthermia. Int J Hyperthermia. 2008; 24: 467–474.
- 27. Pankhurst QA, Connolly J, Jones SK, Dobson J. Applications of magnetic nanoparticles in biomedicine, J Phys D. 2003; 36: R167–R181.
- 28. Rosensweig RE. Heating magnetic fluid with alternating magnetic field. J Magn Magn Mater. 2003; 252: 370–374.
- 29. Kallumadil M, Tada M, Nakagawa T, Abe M, Southern P, Pankhurst QA. Suitability of commercial colloids for magnetic hyperthermia. J Magn Magn Mater. 2009; 321: 1509–1513.
- 30. Krishnan KM. Biomedical Nanomagnetics: A Spin Through Possibilities in Imaging, Diagnostics, and Therapy. IEEE Trans Magn. 2010; 46: 2523–2558.
- 31. Wildeboer RR, Southern P, Pankhurst QA. On the reliable measurement of specific absorption rates and intrinsic loss parameters in magnetic hyperthermia materials. J Phys D Appl Phys. 2014; 47: 495003.
- 32. Wang S-Y, Huang S, Borca DA. Potential sources of errors in measuring and evaluating the specific loss power of magnetic nanoparticles in an alternating magnetic field. IEEE Trans. Magn. 2013; 49: 255–62.
- 33. Kobayashi H, Hirukawa A, Tomitaka A, Yamada T, JeunM, Bae S, Takemura Y. Self-heating property under ac magnetic field and its evaluation by ac/dc hysteresis loops of NiFe₂O₄ nanoparticles. J Appl Phys. 2010; 107: 09B322.
- 34. Nacev A, Weinberg IN, Stepanov PY, Kupfer S, Mair LO, Urdaneta MG, et al. Dynamic Inversion Enables External Magnets To Concentrate Ferromagnetic Rods to a Central Target. Nano Lett. 2015; 15: 359–364.
- 35. Rosalia RA, Cruz LJ, Duikeren S, Tromp AT, Silva AL, Jiskoot W. CD40-targeted dendritic cell delivery of PLGA-nanoparticle vaccines induce potent anti-tumor responses. Biomaterials. 2015; 40: 88–97.
- 36. Lee JH, Jang JT, Choi JS, Moon SH, Noh SH, Kim JW. Exchange-coupled magnetic nanoparticles for efficient heat induction. Nature Nanotechnology 2011; 6: 418–422.
- 37. Tomitaka A, Koshi T, Hatsugai S, Yamada T, Takemura Y. Magnetic characterization of surface-coated magnetic nanoparticles for biomedical application. J Magn Magn Mater. 2011; 323: 1398–1403.
- 38. Dutz S, Kettering M, Hilger I, Müller R, ZeisbergerM.

- Magnetic multicore nanoparticles for hyperthermia—influence of particle immobilization in tumour tissue on magnetic properties. Nanotechnology. 2011; 22: 265102.
- 39. Fortin JP, Gazeau F, Wilhelm C. Intracellular heating of living cells through Néel relaxation of magnetic nanoparticles. Eur Biophys J. 2008; 37: 223–228.
- 40. Corato RD, Espinosa A, Lartigue L, Tharaud M, Chat S, Pellegrino T, et al. Magnetic hyperthermia efficiency in the cellular environment for different nanoparticle designs. Biomaterials. 2014; 35: 6400–6411.
- 41. Ota S, Yamada T, Takemura Y. Magnetization Reversal and Specific Loss Power of Magnetic Nanoparticles in Cellular Environment Evaluated by AC Hysteresis Measurement. J Nanomater. 2015; 2015: 836761.
- 42. Ota S, Yamada T, Takemura Y. Dipole-dipole interaction and its concentration dependence of magnetic fluid evaluated by alternating current hysteresis measurement. J Appl Phys. 2015; 117: 17D713.
- 43. Mehdaoui B, Tan RP, Meffre A, Carrey J, Lachaize S, Chaudret B, Respaud M. Increase of magnetic hyperthermia efficiency due to dipolar interactions in low-anisotropy magnetic nanoparticles: Theoretical and experimental results. Phys Rev B. 2013; 87: 174419.
- 44. Mamiya H, Jeyadevan B. Hyperthermic effects of dissipative structures of magnetic nanoparticles in large alternating magnetic fields. Sci Rep. 2011; 1: 157.
- 45. Carrey J, Mehdaoui B, Respaud M. Simple models for dynamic hysteresis loop calculations of magnetic single-domain nanoparticles: Application to magnetic hyperthermia optimization. J Appl Phys. 2011; 109: 083921.
- 46. Tan RP, Carrey J, Respaud M. Magnetic hyperthermia properties of nanoparticles inside lysosomes using kinetic Monte Carlo simulations: Influence of key parameters and dipolar interactions, and evidence for strong spatial variation of heating power. Phys Rev B. 2014; 90: 214421.